

Hyperspectral imaging in the UV-range – new steps in phenotyping?

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In cooperation with



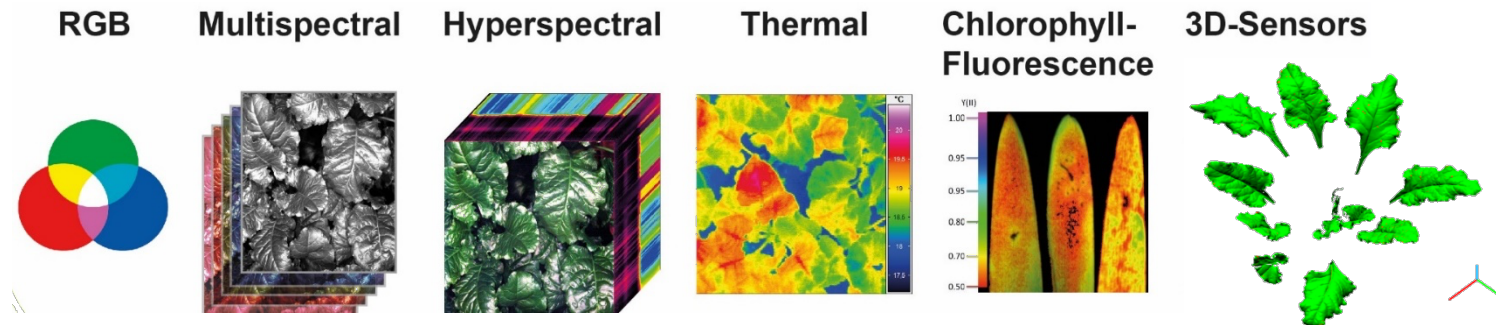
Optical sensors for the detection of diseases

- Host-pathogen interactions differ in their symptom severity, leading to specific spatial patterns and spectral signatures

Mahlein et al. 2012, Plant Meth

- Sensor technologies enable reproducible and objective detection of plant diseases

Rumpf et al. 2010, Comp El Agr



Optical sensors for the detection of diseases: RGB



ISIP Rübenblatt-Scan

ISIP e.V. Tools

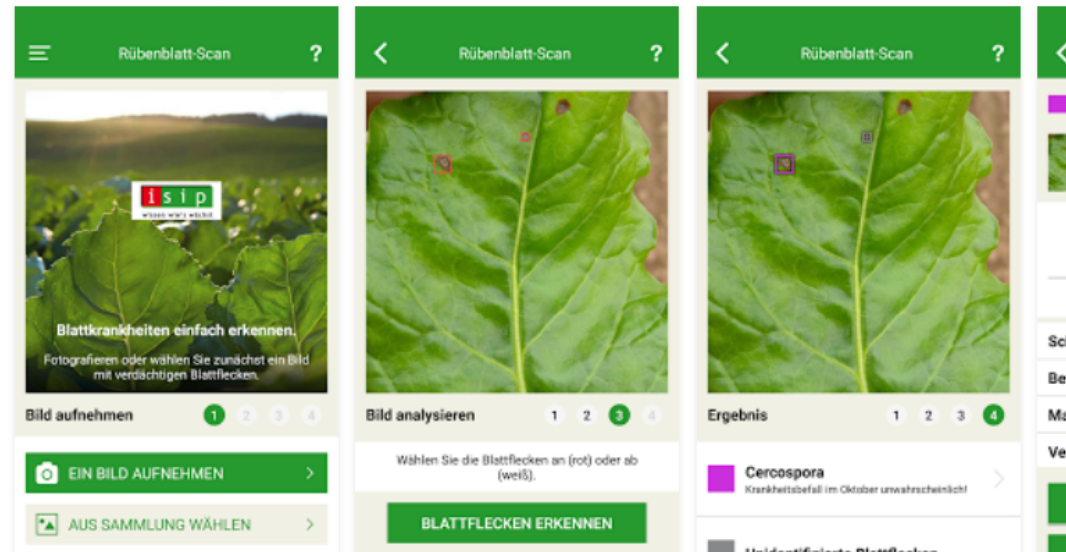
★★★★★ 3

USK ab 0 Jahren

Zur Wunschliste hinzufügen

Installieren

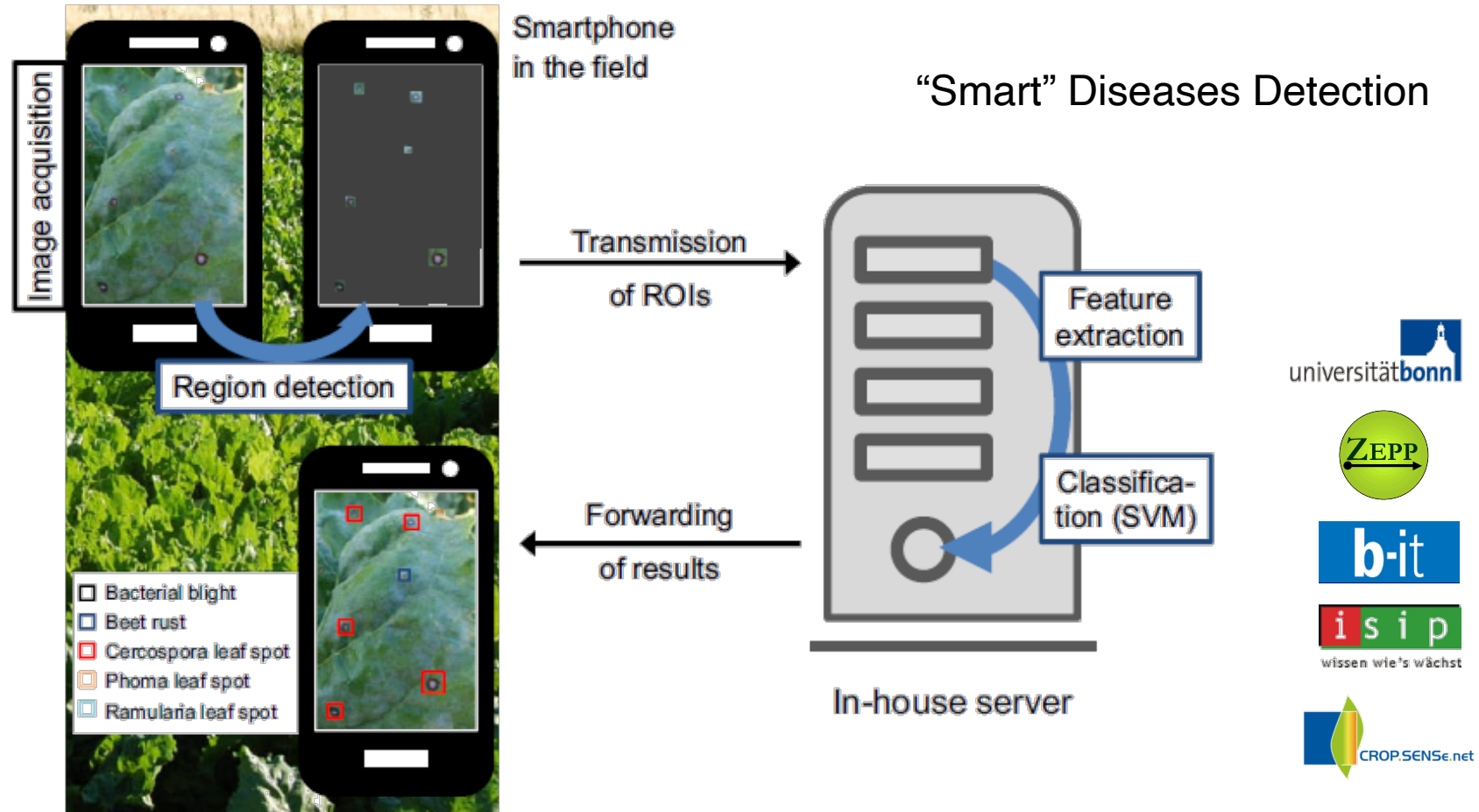
“Smart” Diseases Detection



<https://www.isip.de/isip/servlet/isip-de/apps>

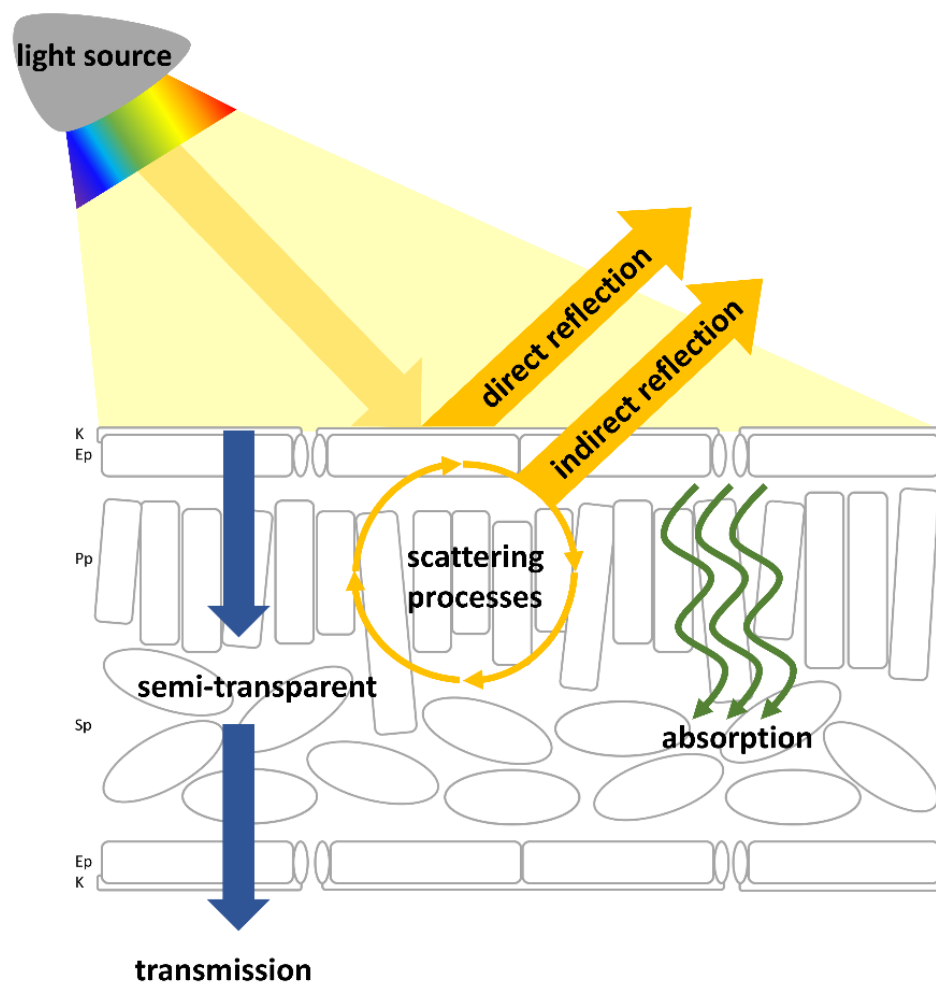


Optical sensors for the detection of diseases: RGB

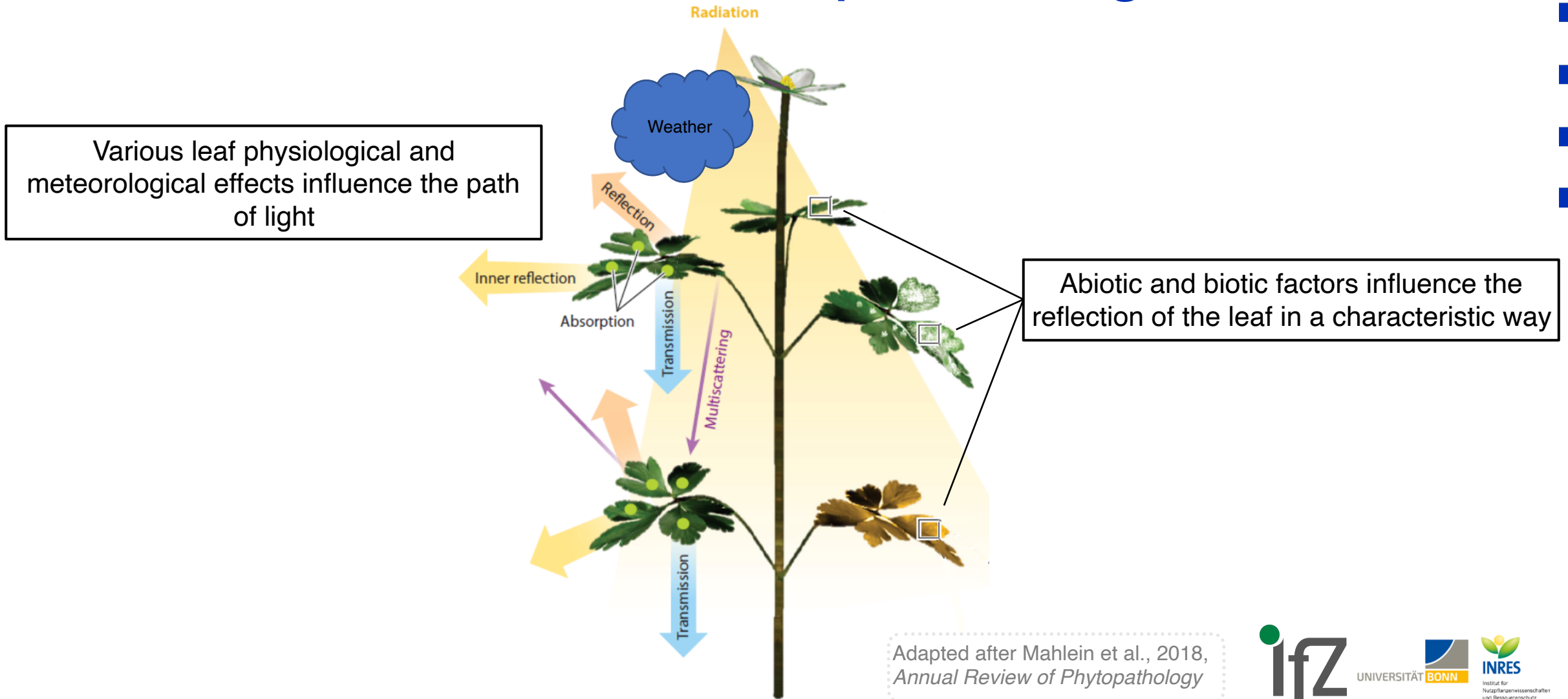


Hallau et al., 2017 *Plant Pathology*

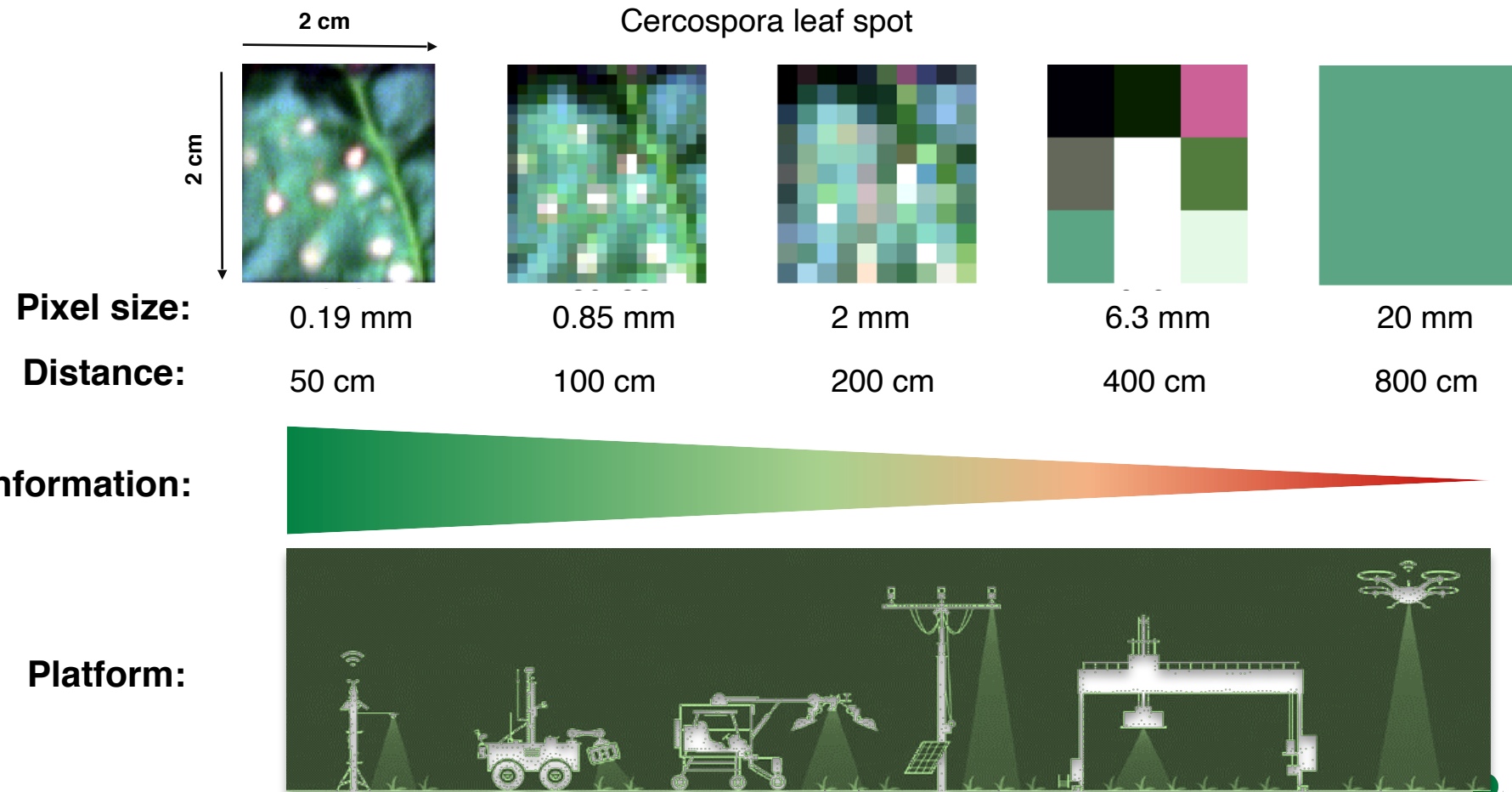
Hyperspectral imaging of plants



Spectral sensors for the detection of diseases – the path of light



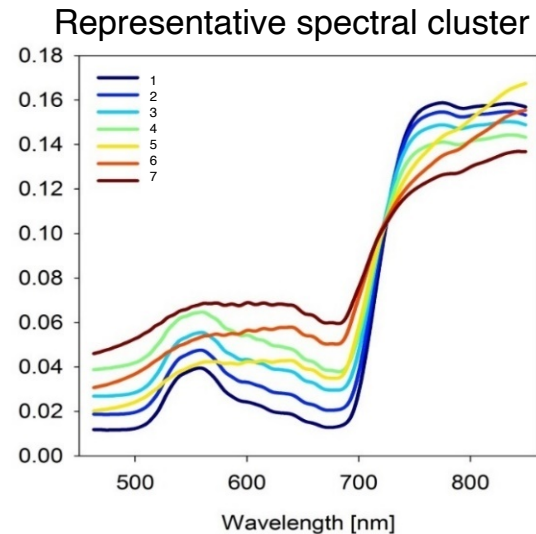
Spectral sensors for disease detection – spatial resolution



Adapted after Shakoor et al., 2017, *Current Opinion in Plant Biology*

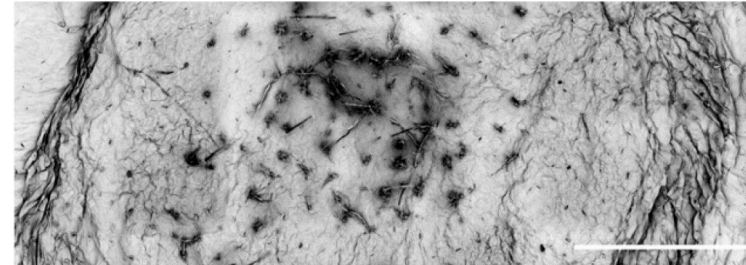
Hyperspectral measurements on different scales: symptom level

Clustering of *Cercospora* symptom-types

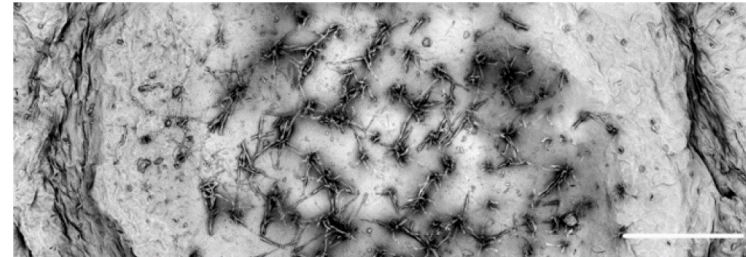


Scanning electron microscopy

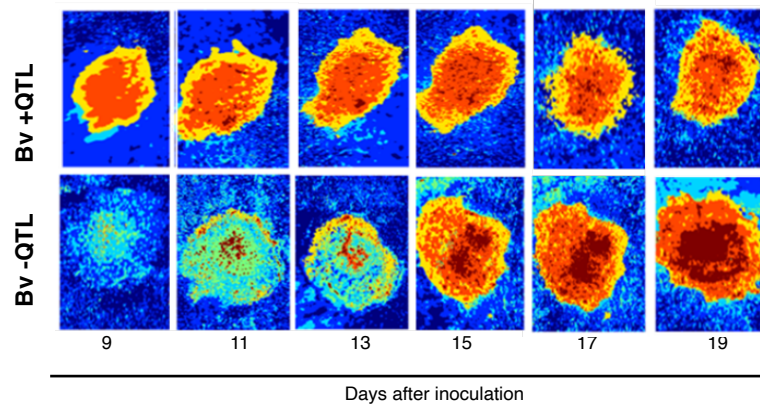
Bv +QTL



Bv -QTL



spectral symptom-phenotypes

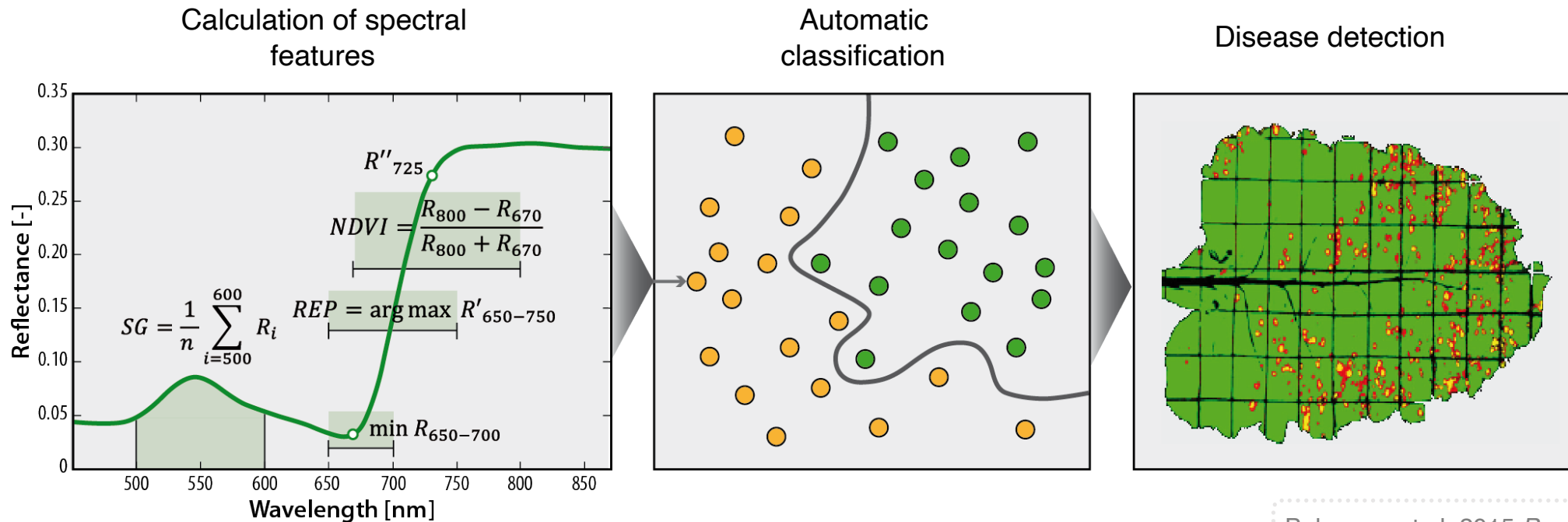


Connection of **spectral phenotypes** and **sporulation intensity**

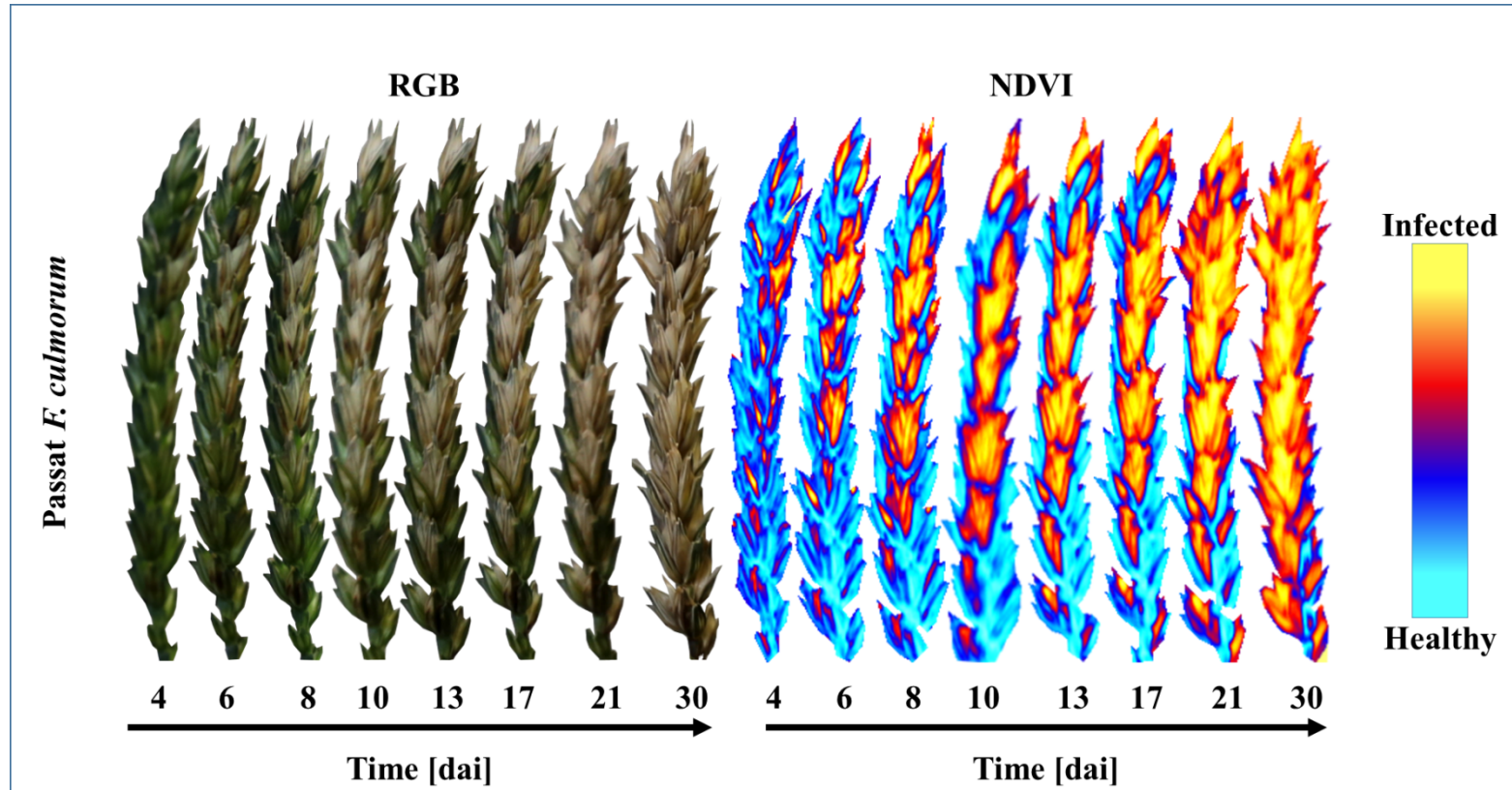
Leucker et al. 2016 *Phytopathology*

Hyperspectral measurements on different scales: leaf level

- Before symptoms are visible?
- Supervised classification using „Support Vector Machines (SVM)“

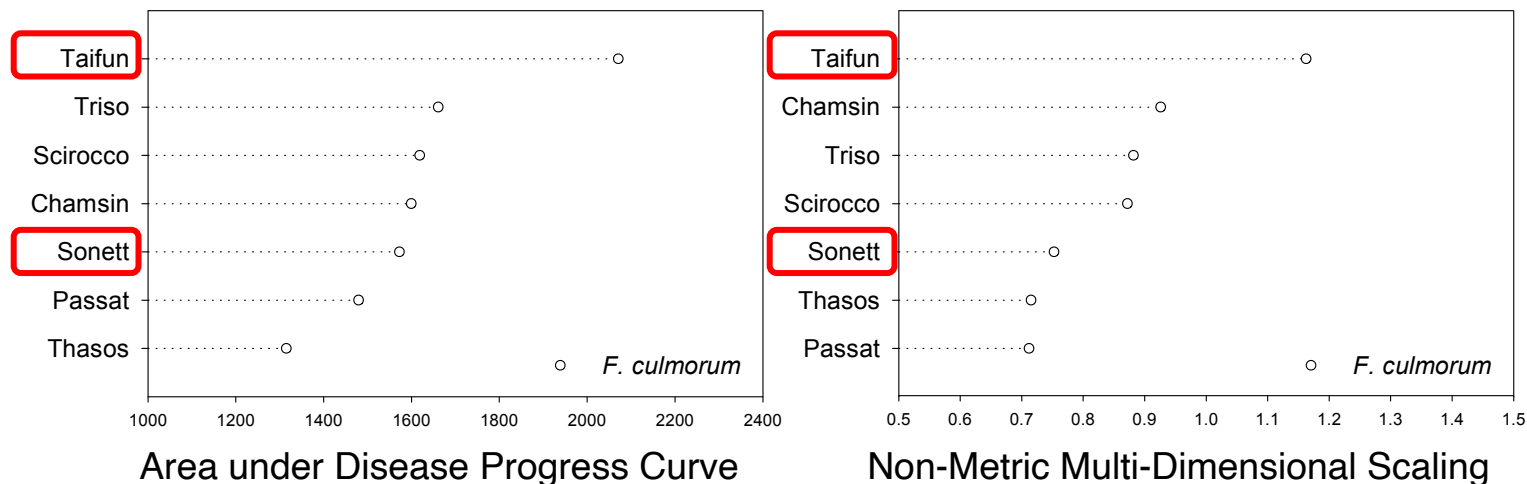
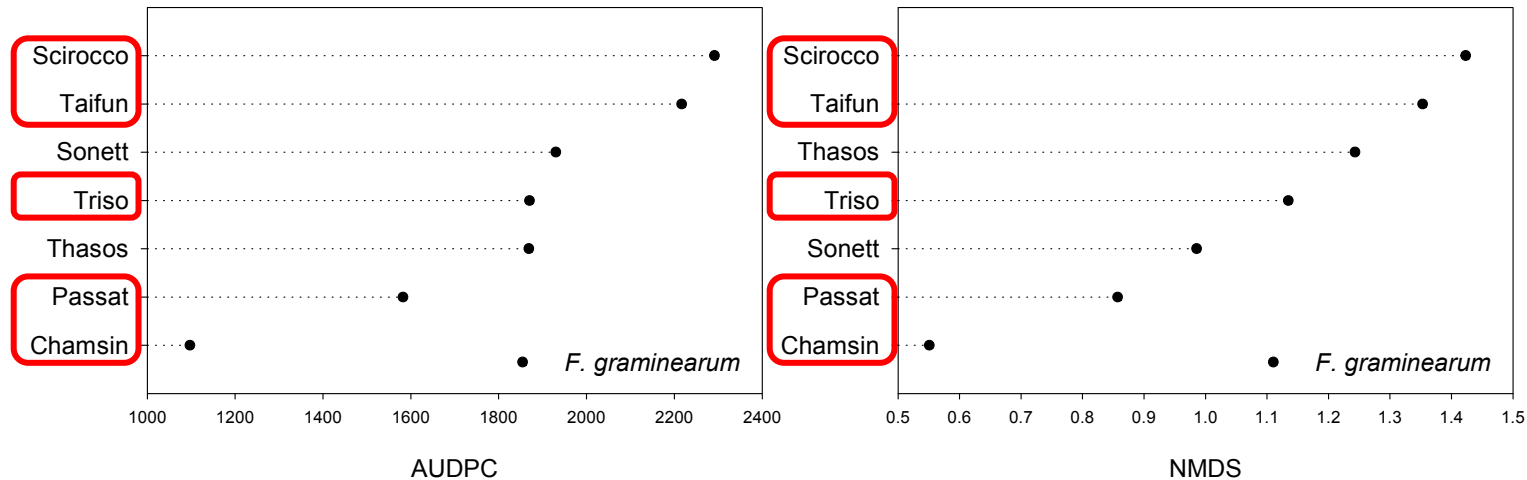


Hyperspectral measurements on different scales: kernel level



Alisaac et al., 2018, *European Journal of Plant Pathology*

Two Ways Variety Ranking



Alisaac et al., 2018, *European Journal of Plant Pathology*

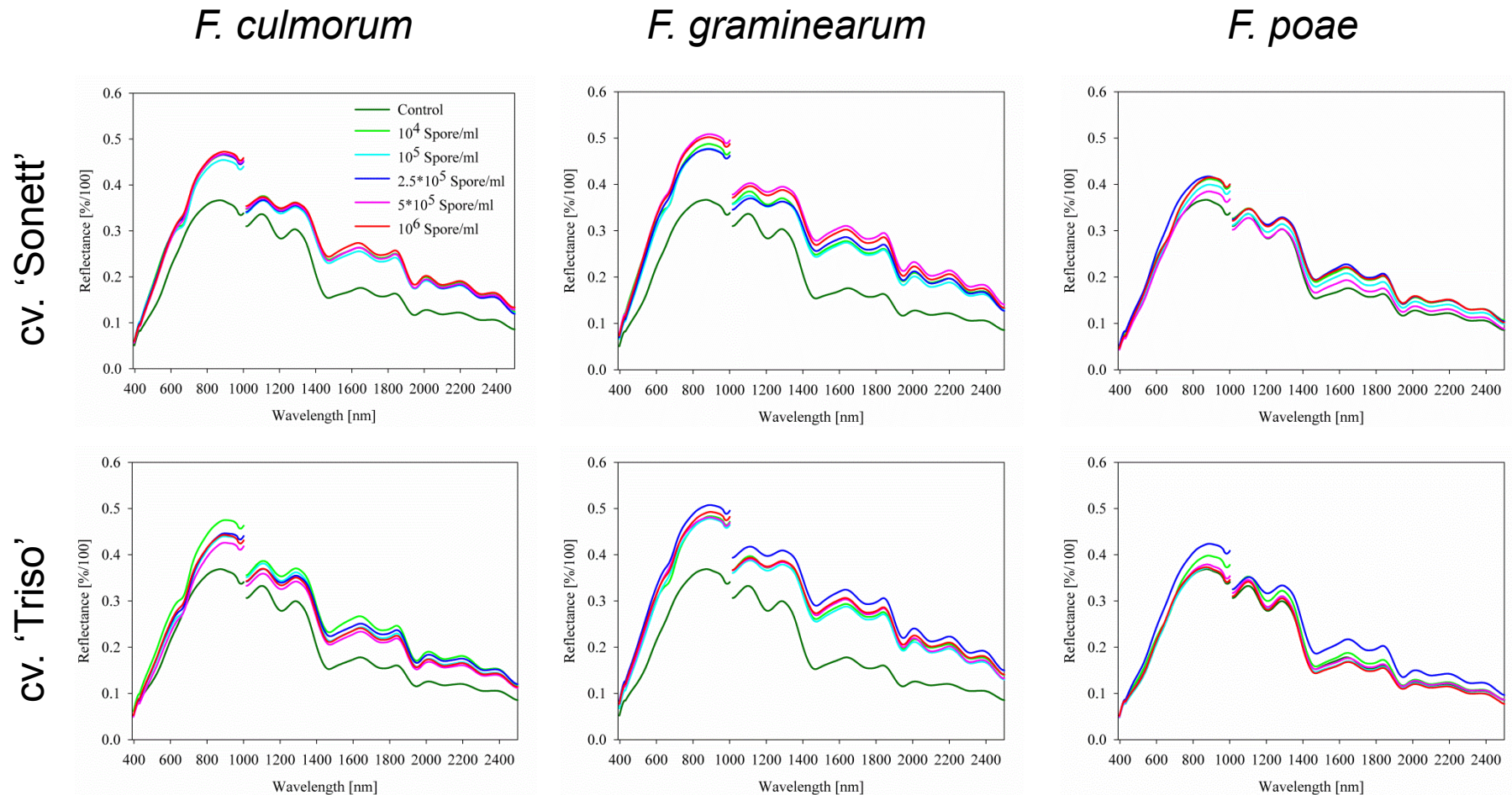
Fusarium infected kernels cv. ('Sonett')



Alisaac et al., 2019, *Toxins*

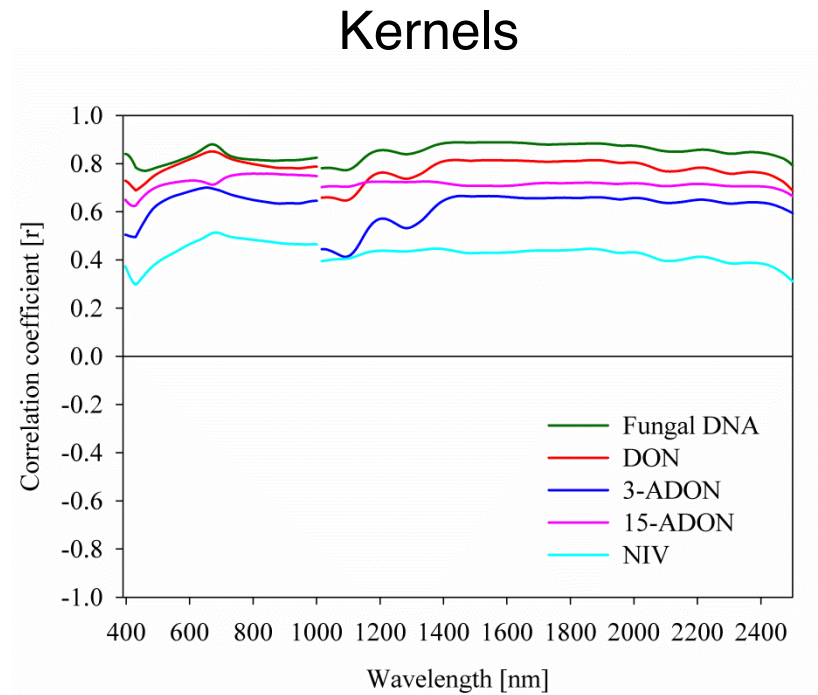
Summer wheat kernels cv. ('Sonett') inoculated with *Fusarium* species (*F. culmorum*, *F. graminearum* and *F. poae*) at different spore densities

Spectral signature of infected kernels

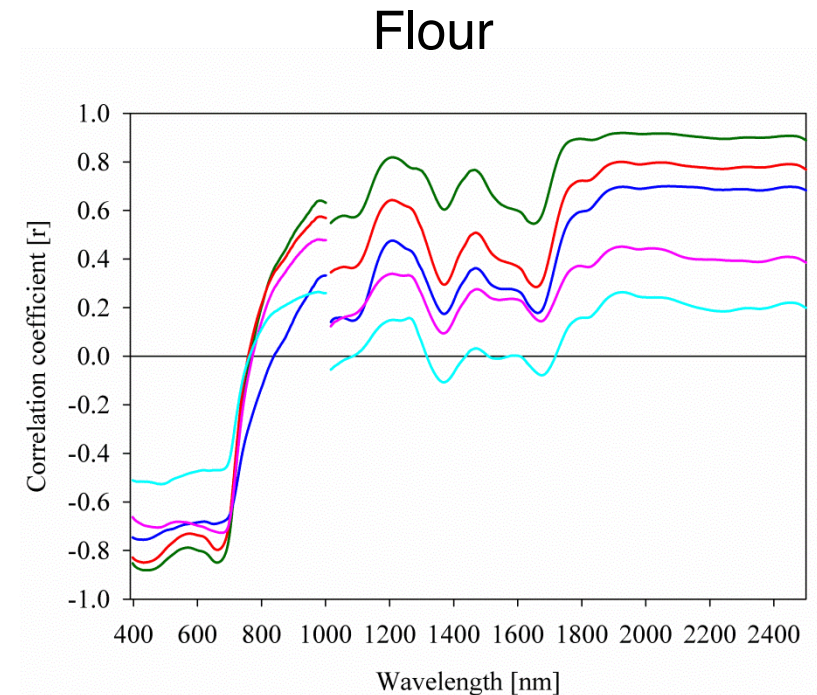


Alisaac et al., 2019, *Toxins*

Correlations between spectral signature in relation with fungal DNA and mycotoxin contents



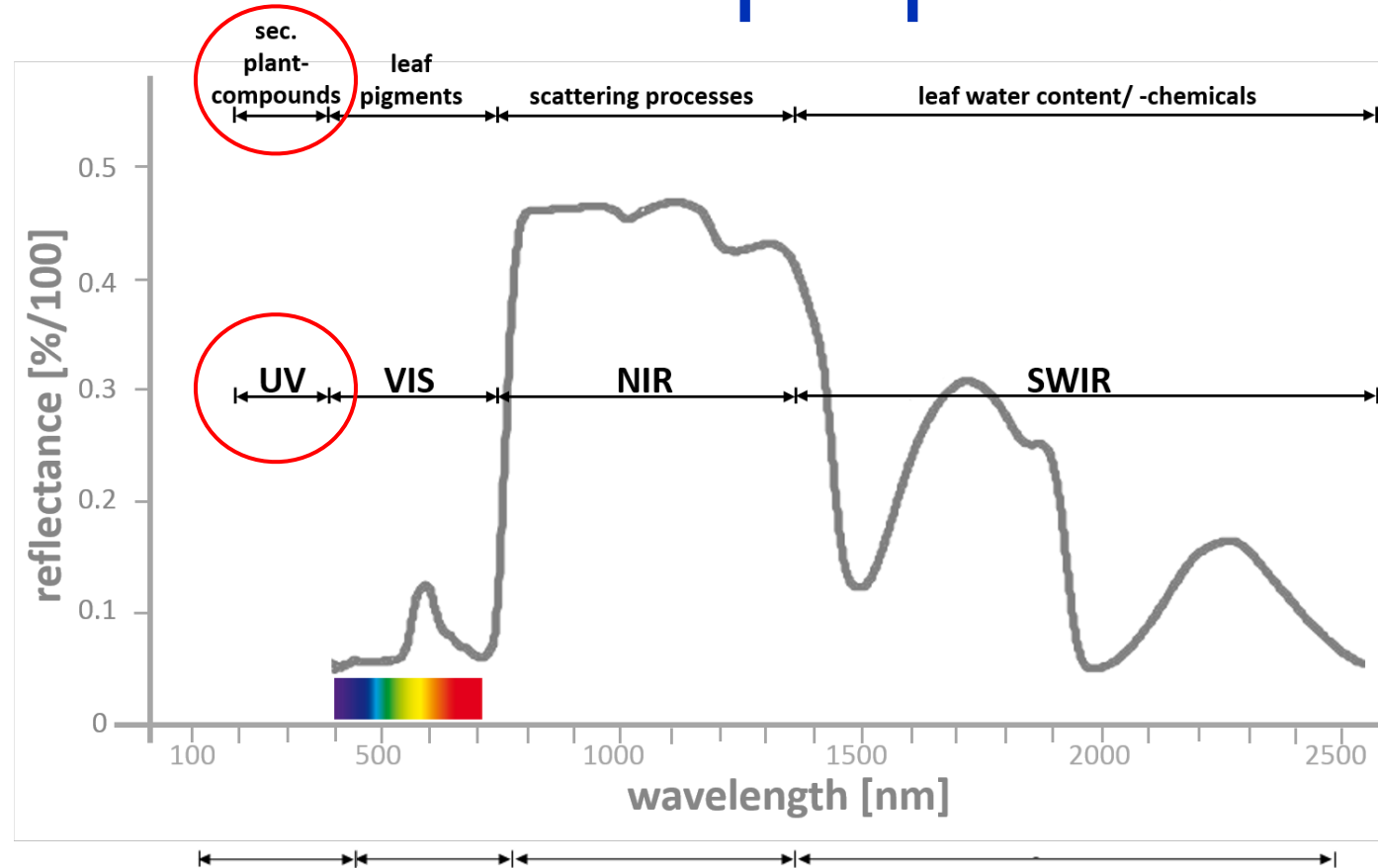
F. graminearum



F. graminearum

Alisaac et al., 2019, *Toxins*

Spectral reflection properties of plants



250-380 nm	380-1030 nm	tomato - <i>Botrytis cinerea</i> (Kong et al. 2014)
?	400-1000 nm	sugar beet – <i>Cercospora beticola</i> (Mahlein et al. 2012)
	400-1000 nm	barley - <i>Erysipales</i> (Kuska et al. 2015)
	400-2500 nm	rape seed - <i>Alternaria</i> (Baranowski et al. 2015)

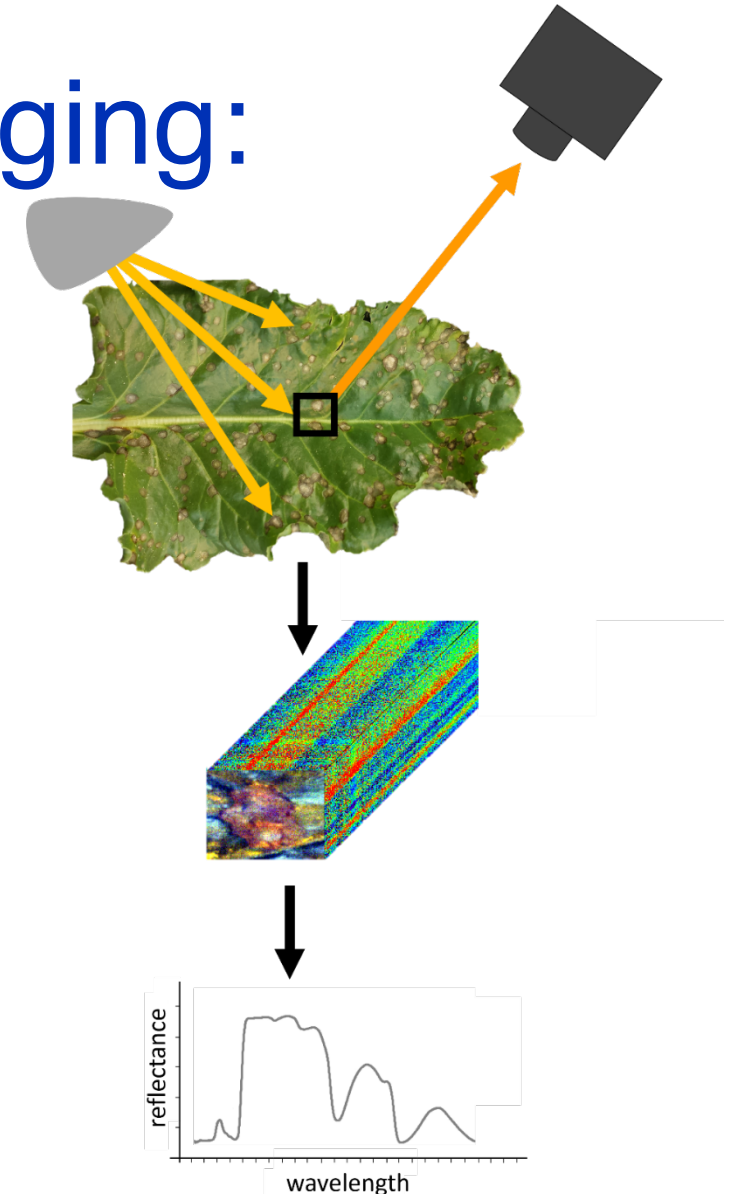
Motivation for UV-imaging:

Early host-pathogen interactions show an influence on plant substances like secondary metabolites (flavones, phenol etc.)

Hypothesis: Changes in the hyperspectral range can be visualized in the UV-range

Requirements to be developed:

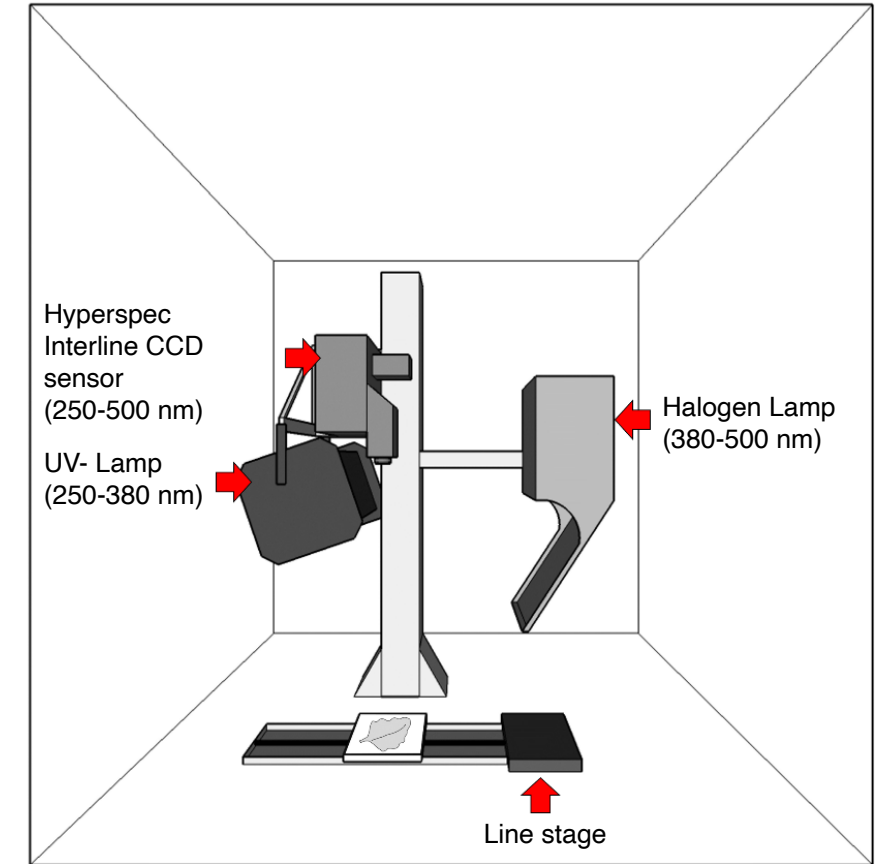
- Homogeneous illumination in the spectral range 250-400 nm
 - Tissue damage to plant material must be avoided
 - Safety aspects during measurements must be taken into account
- Hyperspectral camera with UV-sensor
 - Establishment of measurement protocols
- Development of a processing and evaluation routine
- Biological interpretation of hyperspectral UV signatures



Processing and evaluation of the data

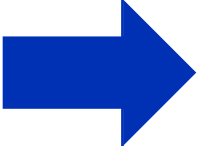
- **Hyperspectral Measurement:**
 - UV-VIS-Interline CCD Sensor (250-500 nm)
 - Spectral resolution 14 nm and focal length 28,3 mm
- **Measurement cabinet:**
 - opaque cabinet
 - 92 cm x 61 cm x 92 cm
- **Illumination:**
 - UV-lamp *Guardian* von UniLux
 - Halogen lamp *21 DC* von TechniQuip
- **Evaluation of data:**
 - ENVI 5.4

Spectral changes of host-pathogen interactions



Phototoxicity of UV-irradiation

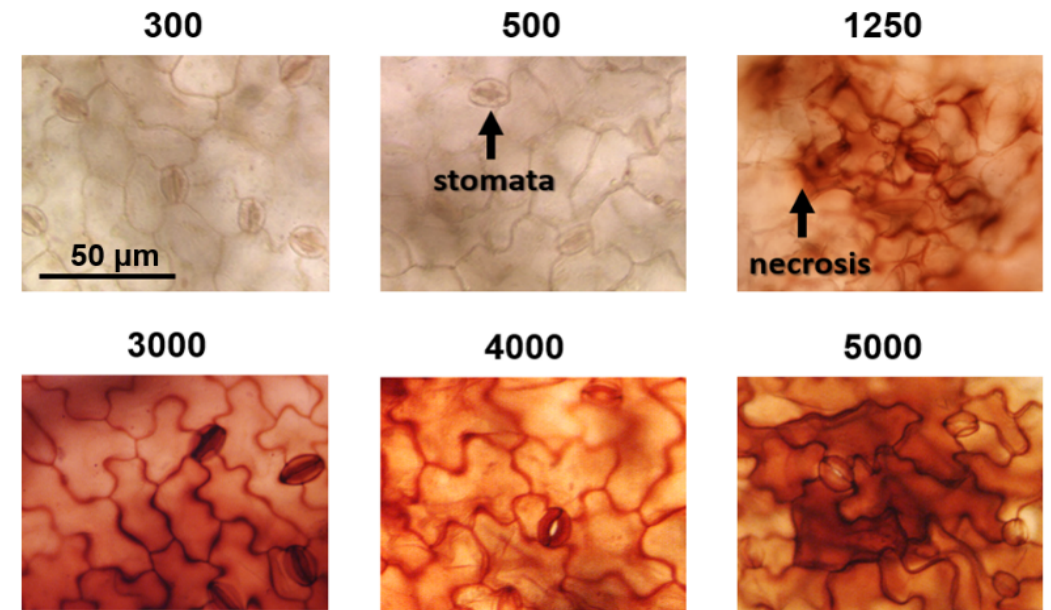
- First measurements of sugar beet leaves revealed tissue damage
- Determination of non-phototoxic UV-intensities
 - Detection of cell death by staining H_2O_2 (DAB-staining after Thordal-Christensen *et al.* 1997)

- 
- **Sugar beet** leaves display no phototoxic effect after illumination with intensities from **300-1250 lux**
 - **Barley leaves** display no phototoxic effect after illumination with intensities from **300-2500 lux**

Brugger et al., 2019, *Remote Sensing*



Tissue damage of sugar beet leaf after illumination with 3500 lux



Microscopic observation of sugar beet leaves after illumination with different intensities

Phototoxicity of UV-irradiation during time-series measurements



8 dai

measuring
every 2 days



16 dai

- Time series measurements of sugar beet leaves showed tissue damage after 5 measurements and illumination with 1250 lux

Time-series measurements can not be conducted with the same sample for sugar beet leaves
Assessment of symptom stages with one sample per measurement

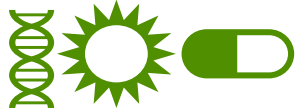
Summary

- Optical sensors can detect, quantify and identify plant diseases
- Smartphone based solutions spread information about plant disease detection
- Application possible on different scales : from cell to field
- Spectral changes also visible from infected kernels
- Hyperspectral measuring protocols in the UV-range established
- Phototoxicity of UV-illumination was evaluated, time-series measurements can not be conducted for sugar beet leaves
 - Symptom stage assessment with one sample per measurement for sugar beet leaves
- Sugar beet leaves inoculated with *C. beticola* and *U. betae* causes changes in the reflectance properties in the UV-range
- Susceptible and resistant barley genotypes lead to different spectral signatures

Plant phenotyping

Aim: accelerate resistance breeding and selection of relevant genotypes

Influenced by: - genotype
- environmental conditions
- management



Size: small plots



Measurement: various traits of genotype, host-pathogen interaction, susceptibility, yield, quality

Technique: RGB imaging, thermal imaging, hyperspectral imaging

Data processing: Delayed data analysis, related to genotyping

Result: Development of new cultivars

Challenges:

- advancement of phenotyping techniques for improving the selection efficiency
- automated monitoring
- cost efficiency

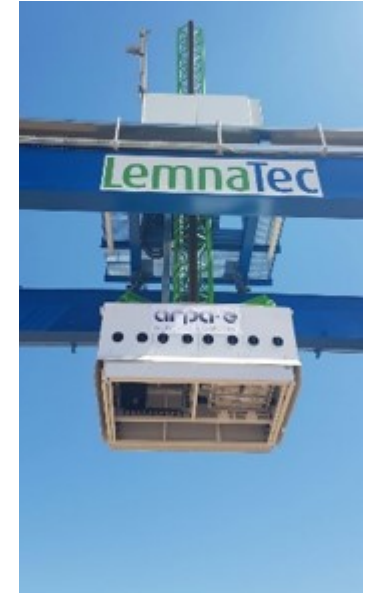
Plant phenotyping examples



Greenhouse Scanalyzer, an automatic plant-to-sensor system in which hyperspectral cameras can be mounted into measurement cabins.



Field Scanalyzer, an automatic sensor-to-plant system in which a camera can be mounted on the probe head.

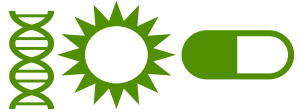


LemnaTec

Precision agriculture

Aim: detect spatial heterogeneity within crop stand

Influenced by: - genotype
- environmental conditions
- management



Size: fields



Measurement: real-time mapping systems of crop, soil and environment variables

Technique: Optical sensors, GPS, imaging platforms, robots, UAVs, satellite images

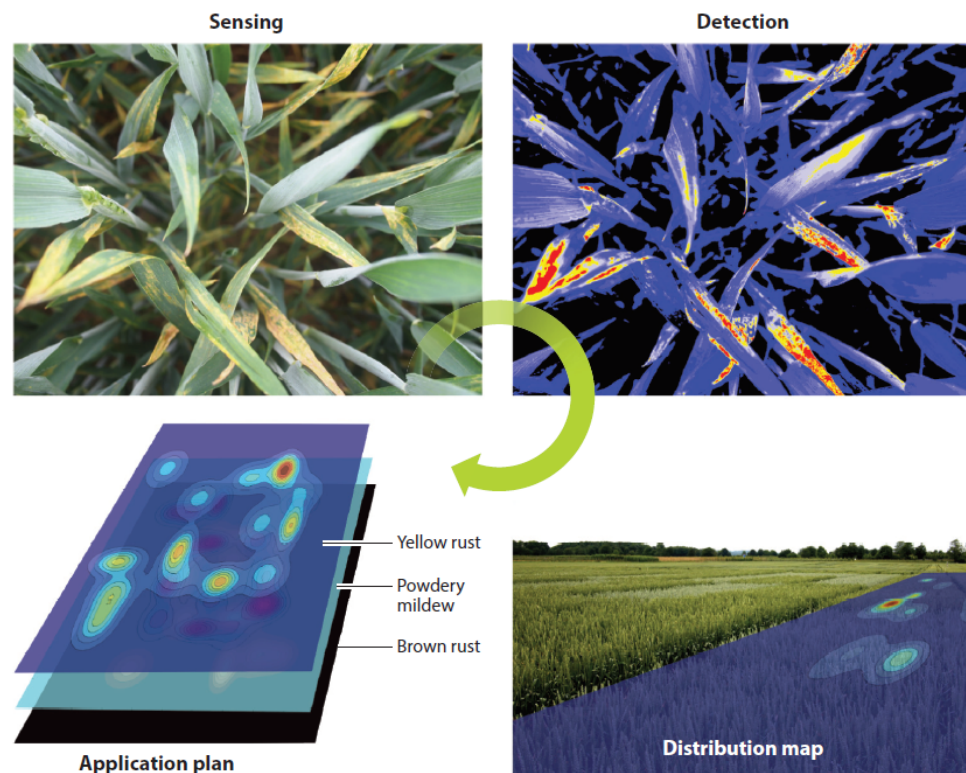
Data processing: Real time data analysis, online, automated data analysis

Result: Decision support for management practice

Challenges:

- quality of the acquired data and data analysis
- automated monitoring
- cost efficiency

Precision agriculture

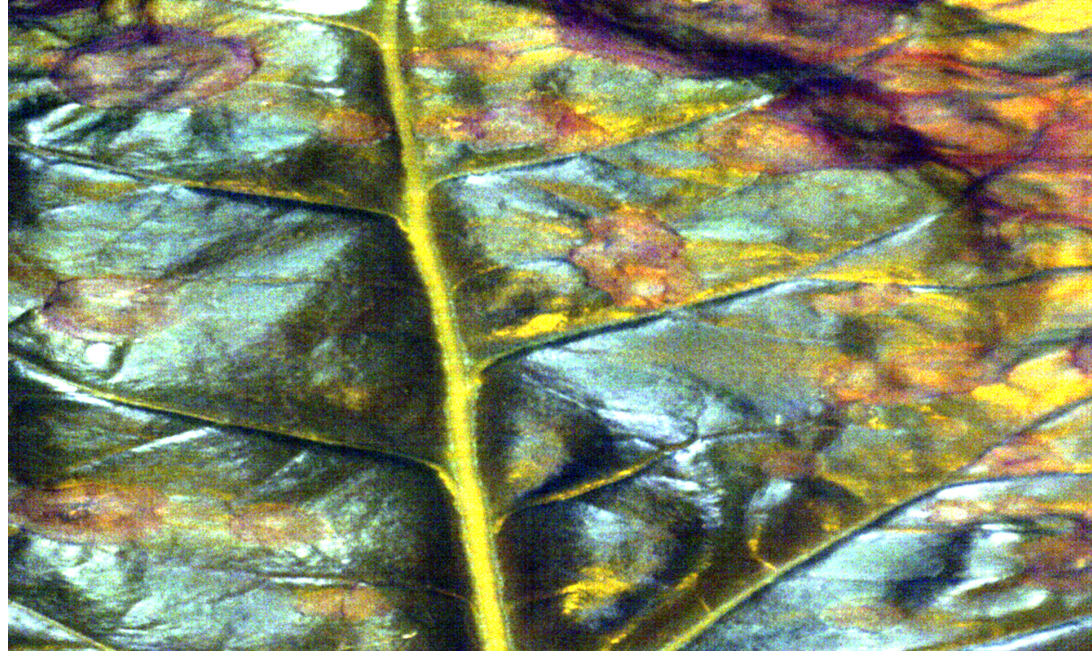


Mahlein et al., 2018, Annual Review of Phytopathology

Outlook

- Research findings have to be transferred into practice
- Implement sensor into plant phenotyping to accelerate resistance breeding and selection of relevant genotypes
- Implement sensor into precision agriculture with the aim to exam spatial heterogeneity
 - Imaging platforms, robots and UAVs
- Data has to be evaluated with machine learning techniques and data mining methods
- Hyperspectral changes will be linked to microscopically and molecular biologically recorded interaction parameter
- Cooperation with project partners for evaluation with data-driven/ “non-supervised” high-throughput methods

Thank you for your attention



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und Landwirtschaft

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